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The absence of AhR in CD4⁺ T cells in patients with acute graft-versus-host disease may be related to insufficient CTCF expression

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Abstract

Background: Acute graft-versus-host disease (aGVHD) is a life-threatening complication of allogeneic hematopoietic stem cell transplantation (allo-HSCT). Accumulating evidence suggests that imbalanced Treg/Th17 ratio accelerates the progression of aGVHD. The aryl hydrocarbon receptor (AhR) is a basic helix-loop-helix transcription factor activated through cognate ligand binding. Current evidence supports that AhR plays a critical regulatory role in the differentiation of Treg and Th17 cells. However, the relationship between AhR and aGVHD remains unclear.

Results: Our results showed that AhR expression was downregulated significantly in CD4⁺ T cells from patients with aGVHD compared with the non-aGVHD group. We also discovered that after activating AhR deficient CD4⁺ T cells, the expression levels of the activation markers-CD40L, CD134 and CD137 and cell proliferation activity were significantly higher than those of AhR-expressing CD4⁺ T cells. Restoring the expression of AhR in aGVHD CD4⁺ T cells resulted in significantly increased percentage of Tregs and associated gene transcripts, including Foxp3, IL-10 and CD39. In contrast, Th17 cell amounts and the transcription of related genes, including ROR γ t, IL-17A and IL-17F, were significantly reduced. We confirmed that CTCF recruited EP300 and TET2 to bind to the AhR promoter region and promoted AhR expression by mediating histone H3K9/K14 hyperacetylation and DNA demethylation in this region. The low expression of CTCF caused histone hypoacetylation and DNA hypermethylation of the AhR promoter, resulting in insufficient expression in aGVHD CD4⁺ T cells.

Conclusions: CTCF is an important inducer of AhR transcription. Insufficient expression of CTCF leads to excessive AhR downregulation, resulting in substantial CD4⁺ T cell activation and Th17/Treg ratio increase, thereby mediating the occurrence of aGVHD.

Keywords: aGVHD, AhR, CTCF, TET2, EP300

Introduction

Although allogeneic hematopoietic stem cell transplantation (allo-HSCT) is considered the exclusive curative treatment in hematopoietic malignancies, acute graft-versus-host disease (aGVHD) is a leading cause of non-relapse mortality following allo-HSCT [1–3]. aGVHD is a complex inflammatory process, in which the immune response is triggered by alloreactive donor T lymphocytes recognizing the host alloantigen. Upon activation, alloreactive donor T cells migrate to target organs,

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primarily the skin, lung, liver and gastrointestinal tract, where they cause tissue damage characteristic of aGVHD [4–6]. Therefore, controlling the excessive activation of T lymphocytes is the key to preventing and treating aGVHD.

The imbalanced ratio of the CD4⁺ T cell subtypes Treg and Th17 cells (Treg/Th17) accelerates the progression of aGVHD [7–9]. Tregs suppress various T cell-associated inflammatory diseases and alleviate GVHD without weakening the graft-versus-leukemia effect [10, 11]. Clinical studies have shown that Treg rate and Foxp3 expression are significantly lower in severe and mild aGVHD cases compared with patients without aGVHD and healthy donors [12, 13]. Th17 cells are relevant to the pathophysiology of aGVHD and significantly contribute to disease severity [14, 15]. Th17 cell rate and ROR γ t expression are significantly higher in severe and mild aGVHD cases than in patients without aGVHD and healthy donors [12]. Therefore, exploring the molecular mechanism of Treg/Th17 imbalance is the key to analyzing the pathogenesis of aGVHD and developing adequate treatment.

The aryl hydrocarbon receptor (AhR) is a basic helix-loop-helix transcription factor activated through cognate ligand binding. It is located in the cytosol of cells as an inactive complex with numerous proteins. Upon ligand binding, the AhR translocates to the nucleus and undergoes conformational changes, recruiting coactivators or corepressors to bind to the promoters of target genes and regulate their expression [16, 17]. Current evidence reveals that the AhR plays a critical role in the regulation of the immune system [18, 19], specifically the differentiation of Treg and Th17 cells [20–23]. AhR activation by 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD), (1'H-indole-3'-carbonyl)-thiazole-4-carboxylic acid methyl ester (ITE), Kynurenine (Kyn), or Laquinimod metabolites increases Foxp3⁺ Treg cells through different mechanisms, including direct transactivation and the induction of epigenetic modifications that control Foxp3 transcription [23–27]. On the other hand, AhR activation by 6-Formylindolo [3,2-b] carbazole (FICZ) boosted Th17 cell differentiation by inducing the expression of RORC [28]. These divergent outcomes following AhR activation have been interpreted as reflecting ligand-intrinsic effects on CD4⁺ T cell differentiation, although no definitive explanation for such differences has been reported.

AhR has been confirmed to inhibit the expression of aGVHD-related inflammatory factors such as IL-1 β , IL-6, and NLRP3 [29, 30]. TCDD was reported to induce regulatory function in CD4⁺ T cells suppressing murine aGVHD in an AhR-dependent mechanism [31]. In addition, studies have found that recipient mice receiving AhR^{-/-} T cells have improved survival and decreased

aGVHD in two different murine allogeneic bone marrow transplant models [32]. Therefore, the relationship between AhR and aGVHD remains unknown, and the expression status of AhR in human aGVHD CD4⁺ T cells is also unclear.

In this study, we demonstrated that the AhR was downregulated in CD4⁺ T cells from patients with aGVHD compared with non-aGVHD cases. To explore the molecular mechanism of AhR downregulation in CD4⁺ T cells in aGVHD, we used bioinformatics and found that the AhR promoter region contains multiple CCCTC binding factor (CTCF) binding sites, as well as multiple CpG islands. It was reported that CTCF binding to target genes leads to DNA demethylation in the occupied region, but the specific mechanism is unclear [33]. Protein–protein interaction analysis showed that CTCF could interact with TET2 and EP300.

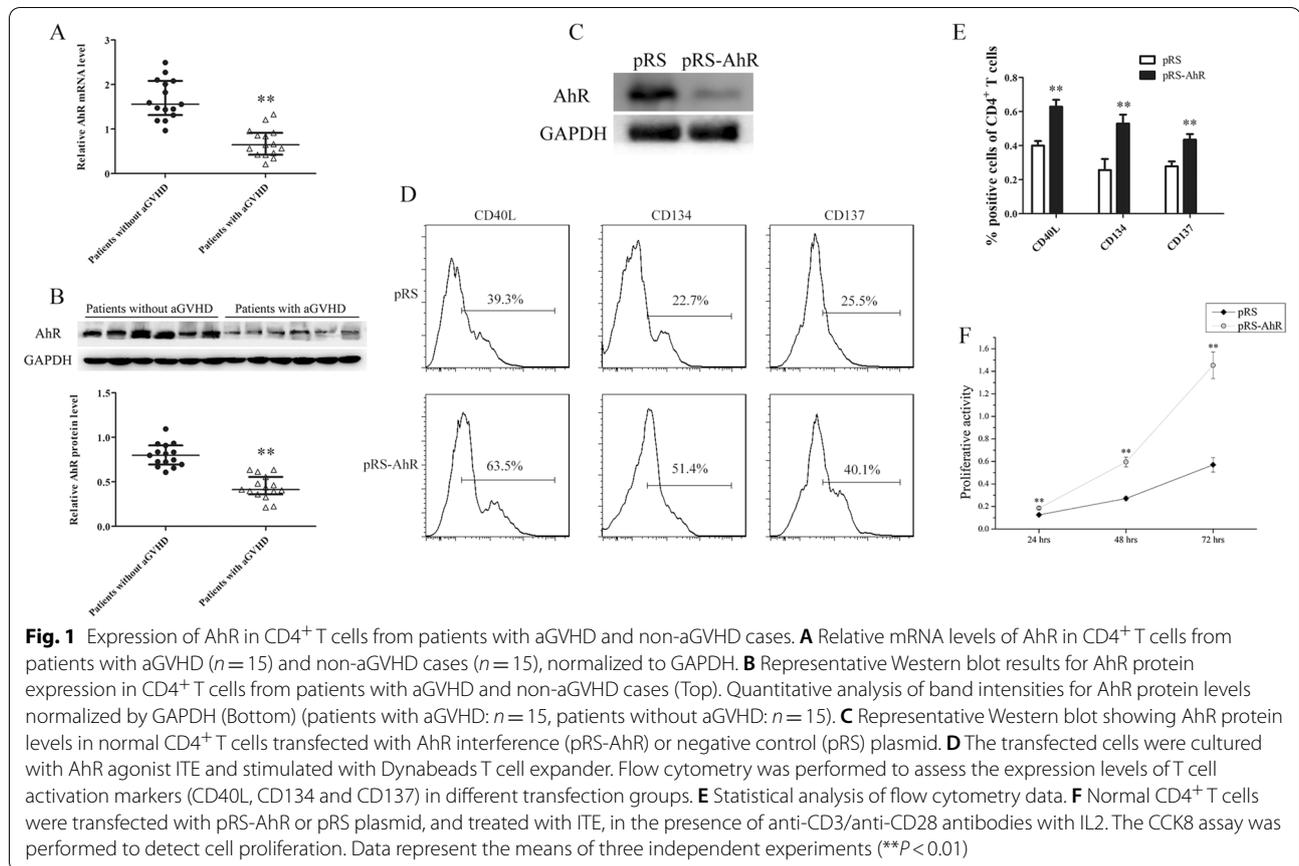
TET2, a member of the ten-eleven translocation (TET) family, can oxidize 5-methylcytosine (5mC) to 5-hydroxymethylcytosine (5hmc), thus promoting DNA demethylation [34]. E1A binding protein p300 (EP300) represents a histone acetyltransferase that regulates transcription via chromatin remodeling and is important in the processes of cell proliferation and differentiation [35]. Histone H3 Lys 9 (H3K9) and H3K14 are common acetylation sites for EP300 modifications and are considered hallmarks of activated gene transcription [36, 37]. Our previous studies confirmed that CTCF expression is reduced in CD4⁺ T cells from aGVHD patients compared with non-aGVHD cases [38]. Therefore, we speculated that low expression of CTCF may reduce the interaction of TET2 with EP300 in the AhR promoter and induce AhR promoter DNA hypermethylation and histone H3K9/K14 hypoacetylation, leading to insufficient AhR expression.

Results

AhR is decreased in CD4⁺ T cells from patients with aGVHD

To investigate the AhR is abnormally expressed in CD4⁺ T cells from GVHD patients, AhR expression levels in CD4⁺ T cells from aGVHD and non-aGVHD patients were examined. Real-time PCR and Western blot showed that AhR expression was downregulated significantly in CD4⁺ T cells from patients with aGVHD compared with the non-aGVHD group (Fig. 1A, B).

To assess the effect of decreased AhR expression on CD4⁺ T cell activation, we transfected the AhR interference (pRS-AhR) or negative control (pRS) plasmid into CD4⁺ T cells from healthy donors. The transfected cells were cultured with AhR agonist ITE and subsequently stimulated with Dynabeads T cell expander (TCR stimulation). Western blot showed that AhR expression in the pRS-AhR group was significantly reduced compared with the pRS group (Fig. 1C). Flow cytometry was performed



to detect the expression levels of T cell activation markers (CD40L, CD134 and CD137) in different transfection groups; compared with the pRS group, the expression levels of CD40L, CD134 and CD137 were significantly higher in the pRS-AhR group (Figures. 1D-E). In addition, to examine the effect of AhR downregulation on CD4⁺ T cell proliferation, normal CD4⁺ T cells were transfected with pRS-AhR and pRS plasmids, respectively, and administered ITE in the presence of anti-CD3/anti-CD28 antibodies with IL2. The CCK8 assay showed that the proliferation of cells transfected with pRS-AhR was significantly higher than that of the pRS group (Fig. 1F). These results indicated that AhR downregulation may be an important reason for the overactivation and proliferation of aGVHD CD4⁺ T cells.

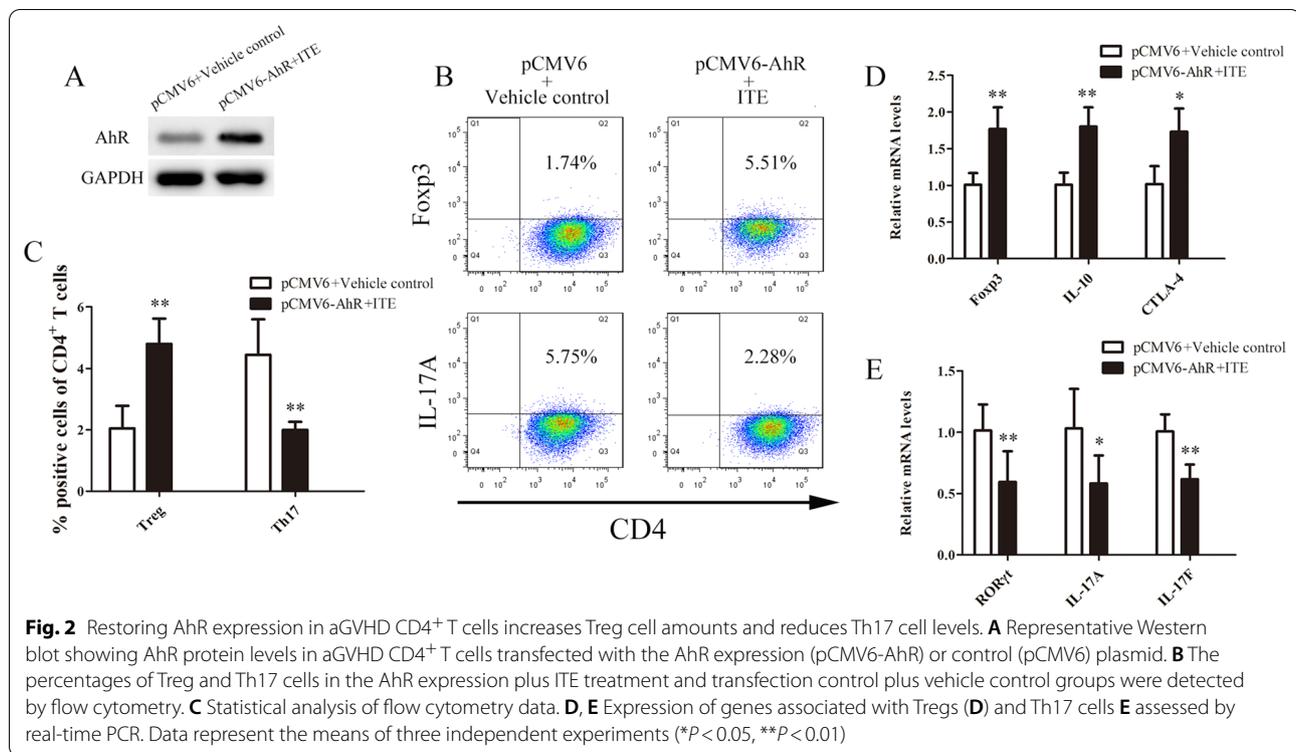
Restoring AhR expression in aGVHD CD4⁺ T cells increases Treg cell amounts and reduces Th17 cell levels

To evaluate the effect of restoring AhR expression in CD4⁺ T cells from patients with aGVHD on Tregs and Th17 cells, we transfected the AhR expression plasmid (pCMV6-AhR) or control (pCMV6) plasmid into aGVHD CD4⁺ T cells. Cells in the AhR expression group were treated with ITE, while those transfected with

pCMV6 were administered the vehicle control. Flow cytometry showed that the percentage of Tregs in CD4⁺ T cells was significantly higher in the AhR expression plus ITE treatment group compared with the transfection control plus vehicle control group (Figures. 2A-C). In contrast, the percentage of Th17 cells was significantly lower in the AhR expression plus ITE treatment group compared with the transfection control plus vehicle control group (Figures. 2A-C). Furthermore, we analyzed the expression levels of genes associated with Tregs and Th17 cells by real-time PCR. AhR expression and ITE treatment led to significantly upregulated Treg-associated gene transcripts, including Foxp3, IL-10 and CTLA-4 (Fig. 2D). In contrast, AhR expression and ITE treatment decreased the expression of Th17-associated gene transcripts, including ROR γ t, IL-17A and IL-17F (Fig. 2E). Collectively, these results demonstrated that restoring AhR expression in aGVHD CD4⁺ T cells could correct Treg/Th17 cell imbalance.

Insufficient CTCF downregulates the AhR in CD4⁺ T cells from patients with aGVHD

To explore the molecular mechanism of AhR downregulation in CD4⁺ T cells from aGVHD patients, we



analyzed the AhR promoter with an online software (<http://jaspar.binf.ku.dk/>) and found that the -204 bp to +611 bp region contained multiple CTCF binding sites (Fig. 3A). ChIP-PCR analysis was used to assess whether CTCF could interact with the AhR promoter in CTCF over-expressed Jurkat cells. Four pairs of real-time PCR primers covering the AhR promoter (-805 to +617 bp) region were used for detection. As shown in Fig. 3B, ChIP-PCR confirmed that CTCF binds in the -204 to +611 bp region of the AhR promoter.

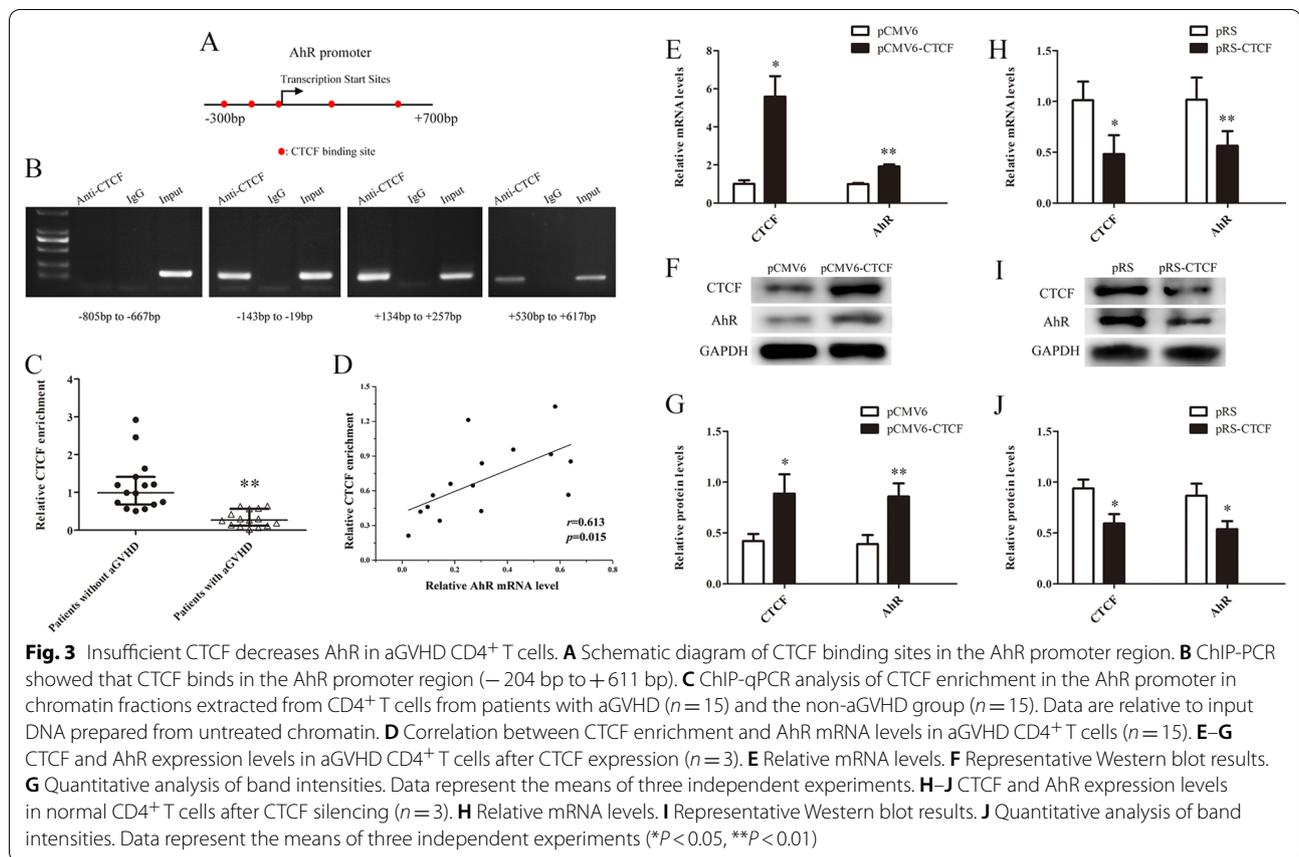
Our previous study confirmed that CTCF expression is reduced in CD4⁺ T cells from aGVHD patients compared with non-aGVHD cases [38]. To further explore the binding levels of CTCF in the AhR promoter region in CD4⁺ T cells from aGVHD patients, we assessed the binding levels of CTCF in the AhR promoter region in CD4⁺ T cells from aGVHD and non-aGVHD patients by ChIP-qPCR using anti-CTCF antibodies. As shown in Fig. 3C, aGVHD patients showed reduced CTCF binding to the AhR promoter compared with the non-aGVHD cases. Correlation analysis indicated an obvious positive correlation between CTCF binding and AhR expression in CD4⁺ T cells from aGVHD patients (Fig. 3D).

To further confirm the promoting effect of CTCF on AhR expression, we transfected the CTCF expression (pCMV6-CTCF) or control (pCMV6) plasmid into aGVHD CD4⁺ T cells and transfected the CTCF

interference (pRS-CTCF) or negative control (pRS) plasmid into normal CD4⁺ T cells. Real-time PCR and Western blot showed that AhR was significantly upregulated in aGVHD CD4⁺ T cells after CTCF expression (Figures 3E-G), and AhR was significantly downregulated in normal CD4⁺ T cells after CTCF silencing (Figures 3H-J). The above findings suggested that CTCF is an important transcription factor inducing AhR expression, and decreased expression of CTCF might be one of the main causes of AhR downregulation in aGVHD CD4⁺ T cells.

CTCF recruits TET2 and EP300 to interact with the AhR promoter region

To further reveal the molecular mechanism of CTCF promoting AhR expression, the hTFtarget database (<http://bioinfo.life.hust.edu.cn/hTFtarget#!/>) was used to predict regulatory factors that may co-regulate AhR expression with CTCF. TET2 and EP300, a methyl cytosine dioxygenase and a histone acetylase, respectively, were predicted to interact with CTCF. Our previous study confirmed that CTCF in CD4⁺ T cells could bind to EP300; so here we only assessed CTCF interaction with TET2 [38]. First, CTCF and TET2 expression plasmids were transfected into Jurkat cells, and co-immunoprecipitation was used to detect whether CTCF could form a complex with TET2. As shown in Fig. 4A, co-immunoprecipitation confirmed that TET2 was co-precipitated



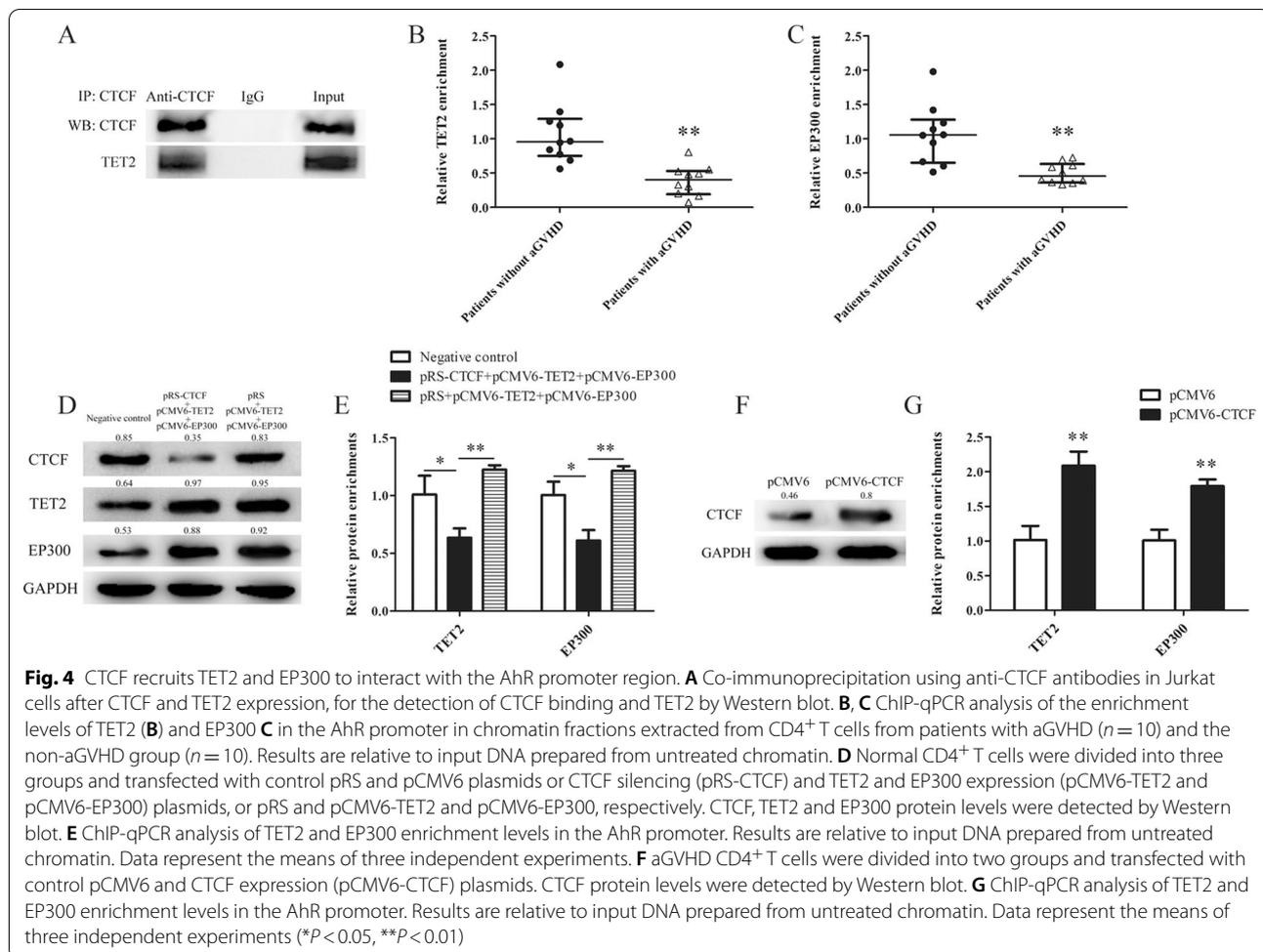
with CTCF. Further, we detected the binding levels of TET2 and EP300 in the AhR promoter region by ChIP-qPCR in CD4⁺ T cells from patients with aGVHD and non-aGVHD cases. As shown in Figures.4B-C, aGVHD patients showed reduced binding levels of TET2 and EP300 at the AhR promoter compared with controls.

To confirm the important role of CTCF in mediating the binding of TET2 and EP300 to the AhR promoter, the CTCF interference and negative control plasmids, respectively, were co-transfected into normal CD4⁺ T cells transfected with TET2 and EP300 expression plasmids (pCMV6-TET2 and pCMV6-EP300), and ChIP-qPCR was used to assess the binding of TET2 and EP300 to the AhR promoter. In the CTCF interference with TET2 and EP300 expression group, the binding levels of TET2 and EP300 in the AhR promoter region were significantly lower compared with those of the scrambled control TET2 and EP300 expression and negative control groups (Figures.4D-E). In addition, we transfected the CTCF expression (pCMV6-CTCF) or control (pCMV6) plasmid into aGVHD CD4⁺ T cells. As shown in Figures.4F-G, TET2 and EP300 binding levels in the AhR promoter region were obviously increased in aGVHD CD4⁺ T cells after CTCF expression. These

results suggested that CTCF is an important transcription factor promoting the binding of TET2 and EP300 to the AhR promoter in CD4⁺ T cells. The reduced binding of TET2 and EP300 in the AhR promoter is associated with insufficient expression of CTCF in aGVHD CD4⁺ T cells.

DNA hypermethylation and histone H3K9/14 hypoacetylation of the AhR promoter in CD4⁺ T cells from patients with aGVHD

TET2 and EP300 are key enzymes that mediate DNA demethylation and histone H3K9/K14 acetylation, respectively. To confirm whether decreased binding of TET2 and EP300 in the AhR promoter causes DNA hypermethylation and histone hypoacetylation in aGVHD CD4⁺ T cells, we detected DNA methylation and histone H3K9/K14 acetylation levels in the AhR promoter region of CD4⁺ T cells from patients with or without aGVHD. BSP showed that the DNA methylation levels of the AhR promoter in CD4⁺ T cells from patients with aGVHD were significantly higher than those of CD4⁺ T cells from non-aGVHD cases (Fig. 5A). ChIP-qPCR showed that the acetylation levels of histone H3K9 and K14 in the AhR promoter in aGVHD CD4⁺



T cells were significantly lower than those of controls (Figures.5B-C). In addition, DNA methylation and histone H3K9/K14 acetylation levels in the AhR promoter region were correlated with AhR expression. The results showed that DNA methylation was negatively correlated with AhR expression (Fig. 5D), while H3K9/K14ac were positively correlated with AhR expression in aGVHD CD4⁺ T cells (Figures.5E-F).

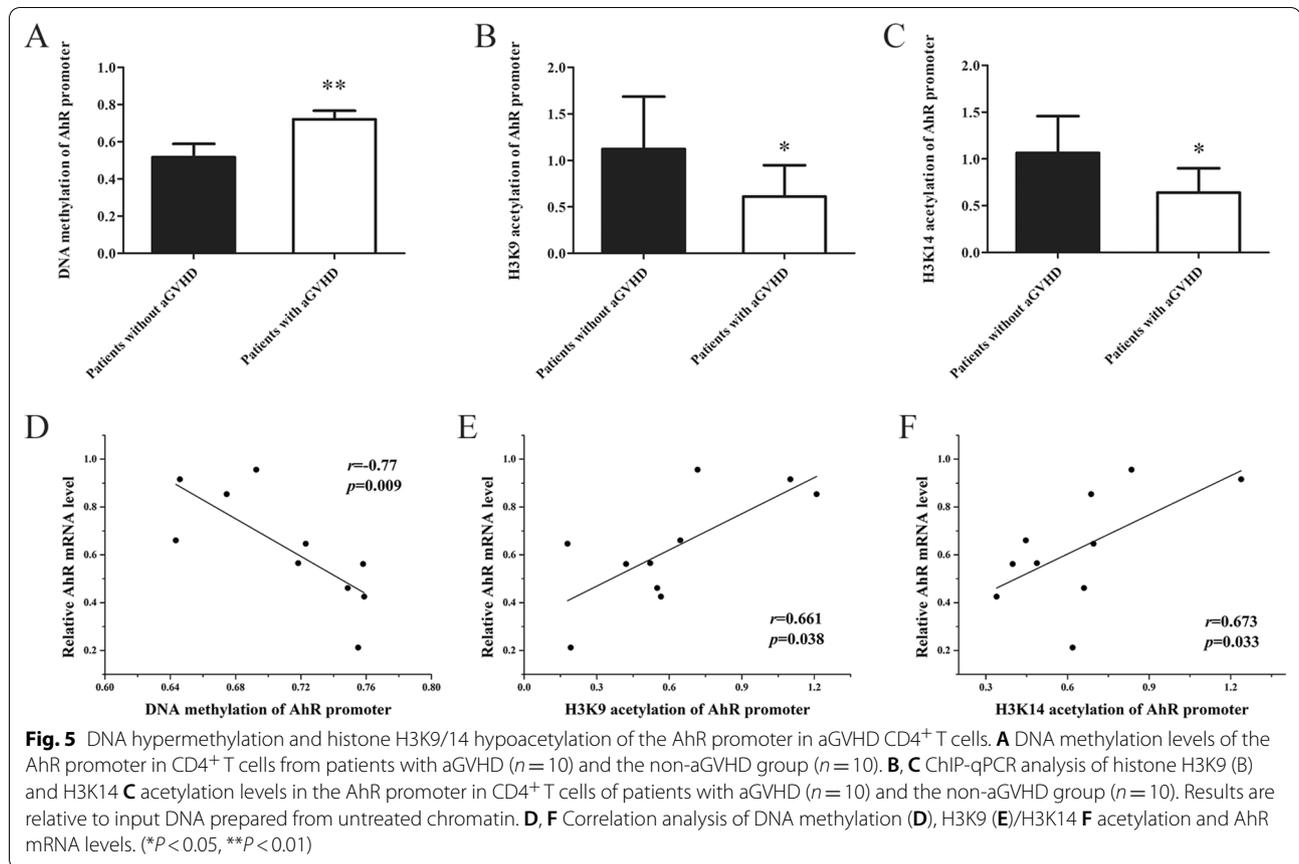
To further confirm the effect of CTCF on DNA methylation and histone H3K9/H3K14 acetylation in the AhR promoter region, DNA methylation and histone H3K9/H3K14 acetylation levels in the AhR promoter region were detected in normal CD4⁺ T cells transfected with the CTCF interference plasmid, or in aGVHD CD4⁺ T cells transfected with the CTCF expression plasmid. We observed increased DNA methylation and decreased histone H3K9/K14ac in the AhR promoter region in normal CD4⁺ T cells after CTCF silencing (Figures.6A-C). Correspondingly, DNA methylation was reduced and H3K9 and K14ac were upregulated significantly in aGVHD CD4⁺ T cells after CTCF expression (Figures.6D-F).

Taken together, these results indicated that CTCF deficiency is an important factor in DNA hypermethylation and histone H3K9/K14 hypoacetylation of the AhR promoter in aGVHD CD4⁺ T cells.

Discussion

CD4⁺ T cells are critical immune effector cells, and alteration in their function has grave consequences on inflammatory responses and immune tolerance [39]. More recently, increasing experimental and epidemiology studies have shown that the AhR plays an important role in immune functions, particularly in the realm of CD4⁺ T cell-mediated inflammation [17, 21].

AhR activation by TCDD promotes the differentiation of Foxp3⁺ Tregs in vivo and in vitro [21, 27]. IL-10-secreting T regulatory type 1 (Tr1) cells are an important subset of CD4⁺ T cells that control excessive inflammation and autoimmunity by inhibiting the functions of antigen-presenting cells and antigen-specific effector T cells [40]. TCDD-mediated AhR activation in CD4⁺ T cells directly changes the expression of genes such as IL-10, CTLA4,

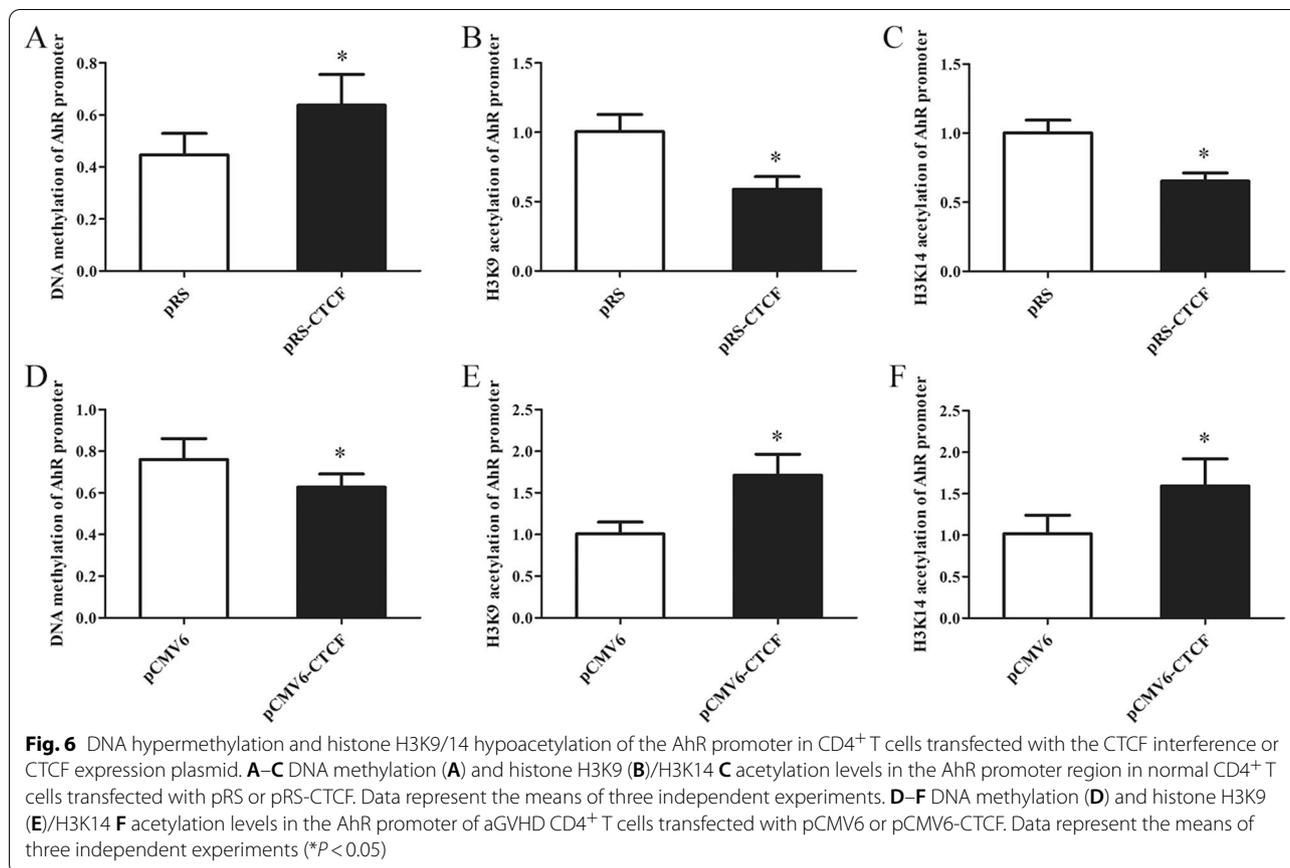


CD39 and GITR, and promotes CD4⁺ T cell differentiation into Tr1 cells [41–44]. In contrast to an increase in Tregs by TCDD, another high-affinity AhR ligand, FICZ, has been reported to enhance Th17 responses and exacerbate immune-mediated diseases in several mouse models [21, 45]. These seemingly contradictory results may be related to the activation strength of the AhR signal. New studies find that low doses of AhR ligands (TCDD, FICZ) did not induce Tregs nor alter the alloresponse, but instead increased the percentage of CD4⁺ T cells that produce IL-17. When administered at high doses, TCDD and FICZ induced Tr1 cells and increased Foxp3⁺ Tregs in conjunction with suppression of the alloresponse [46]. These data indicate that AhR has an impact on both Treg and Th17 differentiation, but Tregs may have a greater demand for activation intensity of AhR signal. Th17 is more sensitive to AhR signal, and a small amount of AhR activation can maintain the differentiation of Th17. We speculated that the attenuation of AhR signal in CD4⁺ T cells has a greater impact on the differentiation of Tregs, resulting in the dominance of Th17 cells.

In this study, AhR expression was significantly down-regulated in CD4⁺ T cells from patients with aGVHD compared with those from non-aGVHD cases. In

addition, our results confirmed that after activating AhR deficient CD4⁺ T cells, the expression levels of the activation markers-CD40L, CD134 and CD137 and cell proliferation activity were significantly higher than those of AhR-expressing CD4⁺ T cells. Studies have shown that AhR endogenous ligands in the plasma of aGVHD patients are significantly reduced, indicating that the AhR signaling pathway is inhibited in aGVHD patients [47, 48]. These results suggest that the normal expression and activation of AhR are important factors in preventing CD4⁺ T cell over activation and proliferation, which may be related to AhR promoting Treg cell differentiation and function. Restoring the expression of AhR in aGVHD CD4⁺ T cells and fully activating it can promote the differentiation of Tregs and induce the expression of functional genes, inhibiting the activation and proliferation of CD4⁺ T cells.

CTCF is a ubiquitously expressed and multifunctional transcription factor implicated in many key cellular processes, including promoter activation and repression, hormone-responsive gene silencing, and genomic imprinting. It is essential and highly conserved from *Drosophila* to mice and humans [49, 50]. Of note, it was recently shown that CTCF may be closely related to Th2



cell and Treg differentiation. Th2 polarization of CTCF-deficient CD4⁺ T cells was blocked, and the transcription levels of IL-4, IL-5 and IL-13 were strongly reduced. In addition, the differentiation of CTCF-deficient CD4⁺ T cells into Tregs was also suppressed, and Foxp3 expression was decreased [51]. CTCF cooperates with Oct-1 to directly interact with the IL-17 locus, regulate Th17 cell differentiation and IL-17 production [52]. These studies showed that CTCF is involved in increasing Tregs and reducing Th17 cells.

Our previous studies have confirmed that CTCF interacts with EP300 to directly bind to the p53 promoter region and promotes p53 expression by upregulating histone H3K9/K14 acetylation levels in this region. The low expression of CTCF in CD4⁺ T cells from aGVHD cases is an important factor leading to histone H3K9/K14 hypoacetylation in the p53 promoter and downregulation p53. Restoring CTCF expression in CD4⁺ T cells from aGVHD patients increases p53 amounts and corrects the imbalance of Th17 cells/Tregs [38]. In the present study, we confirmed that CTCF is an important activator of AhR transcription. It recruited EP300 and TET2 to bind to the AhR promoter region and promoted AhR expression by mediating

histone H3K9/K14 hyperacetylation and DNA demethylation in this region. The low expression of CTCF caused histone hypoacetylation and DNA hypermethylation of the AhR promoter, resulting in insufficient expression in aGVHD CD4⁺ T cells.

EP300 is one of several acetyltransferases that regulate Foxp3 acetylation and functions. Conditional deletion or pharmacologic inhibition of EP300 in Foxp3⁺ Tregs increases T cell receptor-induced apoptosis in Tregs and impairs Tregs' suppressive function as well as peripheral Treg cell induction [53, 54]. TET2 is a key factor that catalyzes the demethylation of the Treg-specific demethylated region (TSDR) of the Foxp3 gene. TET2 mediates TSDR demethylation and activates Foxp3 transcription, driving regulatory T cell differentiation. TET2 deletion leads to Foxp3 hypermethylation, impairs Treg cell differentiation and function, as well as autoimmune disease [55, 56]. Thus, EP300 and TET2 promote the differentiation and functional maintenance of Treg cells by upregulating Foxp3. The current findings provide new insights into the molecular mechanism of EP300 and TET2 regulating the balance of Treg/Th17 cell ratio and participating in immunosuppression.

Conclusions

The present data emphasize the critical role of CTCF in the maintenance of stable AhR expression. Insufficient CTCF expression leads to excessively low expression of AhR, resulting in excessive activation of CD4⁺ T cells and Th17 cell/Treg ratio imbalance, thereby mediating the occurrence of aGVHD.

Materials and methods

Subjects

Totally 55 patients administered allo-HSCT from HLA-identical sibling donors in the Central of Hematopoietic Stem Cell Transplantation of Xiangya Hospital from 2017 to 2021 were included. All subjects provided signed informed consent in accordance with the Declaration of Helsinki. This study was carried out in accordance with the recommendations of international ethical guidelines for biomedical research involving human subjects. The study protocol was approved by the Human Ethics Committee of Xiangya School of Medicine, Central South University. The clinical characteristics of the included patients are shown in Table 1. Conditioning regimens were based on our previous study [57]. aGVHD was assessed based on clinical symptoms in accordance with commonly accepted criteria [58, 59]. We analyzed samples from patients at onset of aGVHD ($n = 55$) and time-matched samples from patients without aGVHD ($n = 55$). Peripheral blood samples were collected as soon as possible after aGVHD diagnosis and before starting the therapy. In addition, we collected normal CD4⁺ T cells from 12 medical professionals recruited in Xiangya Hospital for transfection.

Table 1 Clinical Characteristics of Patients

	No GVHD	aGVHD
Number	55	55
Median age	33	35
Sex (female/male)	25/30	23/32
<i>Diagnosis</i>		
ALL	19	15
AML	29	30
MDS	6	8
CML	1	2
<i>Acute GVHD grade</i>		
1		8
2		25
3		19
4		3
Days to aGVHD onset, median (range)		51 (range: 22–93)

Isolation, culture and transfection of CD4⁺ T cells

Peripheral blood mononuclear cells (PBMCs) were obtained by Ficoll density gradient. CD4⁺ T cells were purified from PBMCs using human CD4 beads (Miltenyi, Bergisch Gladbach, Germany) and cultured in RPMI 1640 (Thermo Fisher Scientific, MA, USA) containing 10% FBS, and 1% penicillin/streptomycin. The isolated CD4⁺ T cells were electrotransfected at 2×10^6 /sample. Human T cell nucleofector Kit and Amaxa nucleofector (Lonza, MD, USA) were used to transfect the gene expression (pCMV6) and interference (pRS) plasmids into CD4⁺ T cells. Briefly, CD4⁺ T cells were mixed with the plasmid in 100 μ l of human T cell nucleofector solution. The mixture was electrotransfected using nucleofector program V-024 in the Amaxa nucleofector. The transfected cells were cultured in RPMI 1640 complete medium for 48 h.

ITE treatment, activation and proliferation of CD4⁺ T cells

(1^H-indole-3[′]-carbonyl)-thiazole-4-carboxylic acid methyl ester (ITE) (MedChemExpress, NJ, USA) was dissolved in DMSO. CD4⁺ T cells from healthy donors were transfected with pRS-AhR and pRS plasmids, respectively, and stimulated with ITE (40 μ M) [60]. Then, cells were activated with Dynabeads T cell expander (anti-CD3/CD28; Invitrogen, CA, USA) for 3 days. The expression levels of T cell activation markers, including CD40L, CD134 and CD137, were detected by flow cytometry. In addition, CD4⁺ T cells were transfected with pRS-AhR and pRS plasmids, respectively, and treated with ITE; cell proliferation was stimulated with Dynabeads T cell expander plus IL-2. After 4 days, the cells were inoculated in 96-well plates. Then, 10 μ L of Cell Counting Kit-8 (CCK-8, MedChemExpress) was added to each well, and the plate was incubated at 37 °C for 1 h in a 5% CO₂ incubator. Absorbance was measured at 450 nm on a microplate reader.

After aGVHD CD4⁺ T cell transfection with AhR expression (pCMV6-AhR) or control (pCMV6) plasmid, cells overexpressing AhR were treated with ITE, while control cells were administered the vehicle control (DMSO). Simultaneously, all transfected cells were stimulated by Dynabeads T cell expander for 3 days. The percentages of Treg and Th17 cells in each group were detected by flow cytometry. The expression levels of genes associated with Treg (Foxp3, IL-10 and CTLA-4) and Th17 (ROR γ t, IL-17A and IL-17F) cells were detected by real-time PCR.

Flow cytometry and intracellular cytokine staining

Flow cytometry was performed to assess the expression levels of the T cell activation markers CD40L, CD134

and CD137. Briefly, the activated cells were suspended in ice-cold FACS buffer (PBS-2% FBS) and incubated with fluorochrome labeled mAbs, including anti-CD40L-PE (eBiosciences, Thermo Fisher, 24–31), anti-CD134-FITC (eBiosciences, ACT-35) and anti-CD137-APC (eBiosciences, 4-1BB) for 20 min at 4 °C. As a control, cells were stained with isotype-control mAbs. Cells were then washed in ice-cold FACS buffer and analyzed on a BD FACScan with the Cell Quest software (BD Biosciences, NJ, USA).

Tregs were detected with anti-CD4-PerCP (eBiosciences, MEM-241) and anti-Foxp3-PE mAbs (eBiosciences, PCH101), while Th17 cells were detected with anti-CD4-PerCP and anti-IL-17A-FITC mAbs (eBiosciences, eBio64DEC17). Briefly, for Tregs, activated cells were first surface-stained with anti-CD4-PerCP mAb. Then, the cells were fixed and permeabilized for intracellular staining with the 4% paraformaldehyde fix solution and Perm/Wash buffer (BD Biosciences, NJ, USA), respectively. After fixation and permeabilization, they were stained with anti-Foxp3-PE and analyzed on a BD FACScan. Meanwhile, Th17 cells were incubated for 4 h with 50 nM PMA (Sigma Aldrich) and 0.5 µg/ml ionomycin (Invitrogen, Thermo Fisher) in the presence of 5 g/mL brefeldin A (eBiosciences) in a tissue culture incubator at 37 °C and 5% CO₂. Cells were then labeled with anti-CD4-PerCP mAbs, fixed and permeabilized, and stained with anti-IL-17A-FITC mAbs. As a control, cells were also stained with isotype-control mAbs.

RNA isolation and real-time PCR

The TRIzol Reagent (Invitrogen, CA, USA) was used to isolate total RNA from CD4⁺ T cells. After DNase treatment, RNA was reverse transcribed with SuperScript II reverse transcriptase (Invitrogen), and then, cDNA was amplified by real-time PCR with fluorescent dye SYBR Green Master Mix (Thermo Fisher Scientific, MA, USA) using an ABI Prism 7500 (Thermo Fisher Scientific). The housekeeping gene human glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as internal standard for sample normalization. Data analysis was performed by the $2^{-\Delta\Delta Ct}$ method; $\Delta\Delta Ct = (Ct_{\text{target gene}} - Ct_{\text{internal control}})_{\text{sample}} - (Ct_{\text{target gene}} - Ct_{\text{internal control}})_{\text{control}}$. The primers are listed in Table 2.

Western blot

CD4⁺ T cells were lysed in ice-cold lysis buffer containing proteinase inhibitors (Thermo Fisher Scientific). Then, the lysate was centrifugated at 12,000 g for 15 min at 4 °C. Protein concentrations were determined with the Bradford protein assay kit (Bio-Rad, CA, USA). Next, equal amounts of protein were resolved by 8% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE),

Table 2 Primer sequences for real-time qPCR

	Forward primer	Reverse primer
AhR	TCCACTTCAGCCACCATC	GGGACTCGGCACAATAAA
CTCF	GAGGCTGCTGTGGACGAT	CAGGCAAAGGTAGGGTGTG
Foxp3	GAGAAGCTGAGTGCCATGCA	AGAGCCCTTGTGCGGATGAT
IL10	TGAGAACAGCTGCACCCACTT	TCGGAGATCTCGAAGCATGTTA
CTLA-4	CAGACCTGCCATGCCAATTTG	GGCCCATATACTTGAATGGAACA
ROR γ t	GCTGGTTAGGATGTGCCG	GGATGCTTTGGCGATGA
IL-17A	CAATCCCACGAAATCCAGGATG	GGTGGAGATCCAAGGTGAGG
IL-17F	TGCTCAAGGAAAGGAAGACA	ATGGTGGATGACAGGGGTG
GAPDH	AAGAGCTACGAGCTGCCTGAC	ATGCCCCAGCGGATGAG

followed by electro-transfer onto polyvinylidene difluoride (PVDF) membranes (Bio-Rad, CA, USA). After blocking at 37 °C with 5% milk in PBS containing 0.1% Tween-20 for 2 h, the membranes were incubated with primary antibodies overnight at 4 °C. The primary antibodies tested included anti-AhR, anti-CTCF, anti-TET2, anti-EP300 and anti-GAPDH (Cell Signaling Technology, MA, USA). After extensive washing, the membranes were incubated with horseradish peroxidase-conjugated secondary antibodies for 2 h at room temperature. The Quantity One software (Bio-Rad, CA, USA) was used to assess immunoreactive bands.

Chromatin immunoprecipitation (ChIP)

ChIP analysis was performed with SimpleChIP[®] Plus Sonication Chromatin IP Kit (Cell Signaling Technology) according to the manufacturer's instructions. Briefly, CD4⁺ T cells were fixed for 10 min with 1% formaldehyde at room temperature. Then, cross-linking was stopped with glycine at 0.125 M. The cells were washed twice with ice-cold PBS and then lysed. Chromatin was sheared into 500–1000 base pair fragments. Next, anti-CTCF, anti-TET2, anti-EP300, anti-H3K9ac, anti-H3K14ac and control rabbit IgG (Cell Signaling Technology), respectively, were added to the lysates for overnight incubation. Immune complexes were precipitated with protein A agarose beads, washed and eluted with 100 µl of TE buffer containing 0.5% SDS and 200 µg/ml proteinase K. Purified DNA was used to amplify the target fragment by PCR or real-time PCR. Primers used were as follows: Forward 1 (–805 to –787):5' CCCTTCACTCCCCCTACA 3' and Reverse 1 (–685 to –667):5'TGGGCCTGCAATAACAT3', Forward 2 (–143 to –121):5' GCCCTCAAGGAAGACGGAATG3' and Reverse 2 (–40 to –19):5' ACCGGCTGAATAGCAGGAGCA3', Forward 3 (+134 to +155):5' GCGGGAAGCACCCTGGATTTA' and Reverse 3 (+236 to +257):5' TAGAATCCTGGCCTGGGTGCG', Forward 4 (+530 to +552):5' CCG

CAGTGGTCCCAGCCTACAC' and Reverse 4 (+598 to +617):5' TCATGGTGCCCAGCCGACG'.

Co-Immunoprecipitation

Nuclear proteins from CD4⁺ T cells were extracted with NE-PER™ Nuclear and Cytoplasmic Extraction Reagents (Thermo Fisher Scientific). Then, anti-CTCF antibodies and control rabbit IgG, respectively, were added to nuclear extracts for overnight incubation at 4 °C. Protein A/G PLUS-agarose beads (Millipore) were added to each IP reaction for 2 h with rotation at 4 °C. Agarose beads were harvested by centrifugation at 3000 g for 2 min. The bound proteins were eluted from the solid support with SDS sample loading buffer. These complexes were analyzed by Western blot using anti-CTCF and anti-TET2 primary antibodies.

Genomic DNA extraction and bisulfite sequencing

Genomic DNA was isolated from CD4⁺ T cells with TIANamp Genomic DNA Kit (TIANGEN, Beijing, China). Bisulfite conversion was performed with EpiTect Bisulfite Kit (Qiagen, CA, USA). Two CpG islands within the AhR promoter region were amplified by PCR. Then, PCR products were subcloned into the pGEM-T vector (Promega, WI, USA). Eight independent clones were sequenced for each of the amplified fragments. Primers used were as follows:5' –GGTGGGGTTTTTAAAGGAAGA-3'(forward1) and 5' –AAACTCCCCAACCCAC TACCTC-3'(reverse1); 5' –GGTATTTTTTTTATTTAA GGGGT-3'(forward2) and 5' –TACAAATAAACTACC TAAACCTAAC-3'(reverse2);

Statistical analysis

Statistics were performed using SPSS 22.0 software. Continuous variables with normal distribution were expressed as mean ± SD; non-normally distributed variables were reported as medians with interquartile ranges. Normally distributed continuous variables were compared by independent samples *t* test. Non-normally distributed variables were compared by Mann–Whitney *U* test. Correlations were analyzed using Spearman's correlation coefficients. Statistical significance was defined by *P* < 0.05.

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Author contributions

CZ, TTC, and YJX contributed to the design and planning of the experiments. XM, YL and JH provided samples. CZ, TTC, XM, XC, YL, and SYW conducted the laboratory experimental work. CZ, TTC, XM, JH, XC, and YJX contributed to the reporting of findings and writing of the manuscript. All authors critically revised the manuscript and gave final approval of the version to be submitted. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets supporting the conclusions of this article are included within the article. All other datasets used and analyzed during the study are available from the corresponding author on reasonable request.

Declarations

Competing interests

The authors declare that there are no conflicts of interest regarding the publication of this article.

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References

- Martin PJ, Rizzo JD, Wingard JR, Ballen K, Curtin PT, Cutler C, Litzow MR, Nieto Y, Savani BN, Schriber JR, Shaughnessy PJ, Wall DA, Carpenter PA. First- and second-line systemic treatment of acute graft-versus-host disease: recommendations of the American society of blood and marrow transplantation. *Biol Blood Marrow Transpl J Am Soc Blood Marrow Transpl.* 2012;18:1150–63.
- Wolff D, Ayuk F, Elmaagacli A, Bertz H, Lawitschka A, Schleuning M, Meyer RG, Gerbitz A, Hilgendorf I, Hildebrandt GC, Edinger M, Klein S, Halter J, Mousset S, Holler E, Greinix HT. Current practice in diagnosis and treatment of acute graft-versus-host disease: results from a survey among German–Austrian–Swiss hematopoietic stem cell transplant centers. *Biol Blood Marrow Transpl J Am Soc Blood Marrow Transpl.* 2013;19:767–76.
- Chen Y, Xu Y, Fu G, Liu Y, Peng J, Fu B, Yuan X, Xin H, Zhu Y, He Q, Wu D, Shu Y, Li X, Zhao X, Chen F. Allogeneic hematopoietic stem cell transplantation for patients with acute leukemia. *Chin J Cancer Res.* 2013;25:389–96.
- Zeiser R, Socie G, Blazar BR. Pathogenesis of acute graft-versus-host disease: from intestinal microbiota alterations to donor T cell activation. *Br J Haematol.* 2016;175:191–207.
- Zitler NC, Snyder K, Meng X, Taylor PA, Efebera YA, Devine SM, Blazar BR, Garzon R, Ranganathan P. MicroRNA-155 modulates acute graft-versus-host disease by impacting T Cell expansion migration, and effector function. *J Immunol.* 2018;200:4170–9.
- Xu YJ, Li L, Chen Y, Fu B, Wu DS, Li XL, Zhao XL, Chen FP. Role of HMGB1 in regulation of STAT3 expression in CD4(+) T cells from patients with aGVHD after allogeneic hematopoietic stem cell transplantation. *Clin Immunol (Orlando, Fla).* 2015;161:278–83.
- Han L, Jin H, Zhou L, Zhang X, Fan Z, Dai M, Lin Q, Huang F, Xuan L, Zhang H, Liu Q. Intestinal microbiota at engraftment influence acute graft-versus-host disease via the Treg/Th17 balance in Allo-HSCT recipients. *Front Immunol.* 2018;9:669.
- Di Ianni M, Falzetti F, Carotti A, Terenzi A, Castellino F, Bonifacio E, Del Papa B, Zei T, Ostini RI, Cecchini D, Aloisi T, Perruccio K, Ruggeri L, Balucani C, Pierini A, Sportoletti P, Aristei C, Falini B, Reisner Y, Velardi A, Aversa F, Martelli MF. Tregs prevent GVHD and promote immune reconstitution in HLA-haploidentical transplantation. *Blood.* 2011;117:3921–8.
- Perkey E, Maillard I. New insights into graft-versus-host disease and graft rejection. *Annu Rev Pathol.* 2018;13:219–45.
- Lourenco EV, La Cava A. Natural regulatory T cells in autoimmunity. *Autoimmunity.* 2011;44:33–42.

11. Kingsley CI, Karim M, Bushell AR, Wood KJ. CD25+CD4+ regulatory T cells prevent graft rejection: CTLA-4- and IL-10-dependent immunoregulation of alloresponses. *J Immunol.* 2002;168:1080–6.
12. Liu Y, Cai Y, Dai L, Chen G, Ma X, Wang Y, Xu T, Jin S, Wu X, Qiu H, Tang X, Li C, Sun A, Wu D, Liu H. The expression of Th17-associated cytokines in human acute graft-versus-host disease. *Biol Blood Marrow Transpl.* 2013;19:1421–9.
13. Fujioka T, Tamaki H, Ikegame K, Yoshihara S, Taniguchi K, Kaida K, Kato R, Inoue T, Nakata J, Ishii S, Soma T, Okada M, Ogawa H. Frequency of CD4(+)FOXP3(+) regulatory T-cells at early stages after HLA-mismatched allogeneic hematopoietic SCT predicts the incidence of acute GVHD. *Bone Marrow Transpl.* 2013;48:859–64.
14. Delens L, Ehx G, Somja J, Vrancken L, Belle L, Seidel L, Grégoire C, Fransolet G, Ritacco C, Hannon M, Dubois S, Beguin Y, Baron F, Servais S. In vitro Th17-polarized human CD4(+) T cells exacerbate xenogeneic graft-versus-host disease. *Biol Blood Marrow Transpl.* 2019;25:204–15.
15. Malard F, Gaugler B, Lamarthee B, Mohty M. Translational opportunities for targeting the Th17 axis in acute graft-vs.-host disease. *Mucosal Immunol.* 2016;9:299–308.
16. Shinde R, Hezaveh K, Halaby MJ, Kloetgen A, Chakravarthy A, da Silva Medina T, Deol R, Manion KP, Baglaenko Y, Eldh M, Lamorte S, Wallace D, Chodiseti SB, Ravishankar B, Liu H, Chaudhary K, Munn DH, Tsigiris A, Madaio M, Gabrielsson S, Touma Z, Wither J, De Carvalho DD, McGaha TL. Apoptotic cell-induced AhR activity is required for immunological tolerance and suppression of systemic lupus erythematosus in mice and humans. *Nat Immunol.* 2018;19:571–82.
17. Shinde R, McGaha TL. The aryl hydrocarbon receptor: connecting immunity to the microenvironment. *Trends Immunol.* 2018;39:1005–20.
18. Stevens EA, Mezrich JD, Bradfield CA. The aryl hydrocarbon receptor: a perspective on potential roles in the immune system. *Immunology.* 2009;127:299–311.
19. Gutiérrez-Vázquez C, Quintana FJ. Regulation of the immune response by the aryl hydrocarbon receptor. *Immunity.* 2018;48:19–33.
20. Duarte JH, Di Meglio P, Hirota K, Ahlfors H, Stockinger B. Differential influences of the aryl hydrocarbon receptor on Th17 mediated responses in vitro and in vivo. *PLoS ONE.* 2013;8: e79819.
21. Quintana FJ, Basso AS, Iglesias AH, Korn T, Farez MF, Bettelli E, Caccamo M, Oukka M, Weiner HL. Control of T(reg) and T(H)17 cell differentiation by the aryl hydrocarbon receptor. *Nature.* 2008;453:65–71.
22. Abdulla OA, Neamah W, Sultan M, Chatterjee S, Singh N, Nagarkatti M, Nagarkatti P. AhR ligands differentially regulate miRNA-132 which targets HMGB1 and to control the differentiation of tregs and Th-17 cells during delayed-type hypersensitivity response. *Front Immunol.* 2021;12: 635903.
23. Kerkvliet NI, Steppan LB, Vorachek W, Oda S, Farrer D, Wong CP, Pham D, Mourich DV. Activation of aryl hydrocarbon receptor by TCDD prevents diabetes in NOD mice and increases Foxp3+ T cells in pancreatic lymph nodes. *Immunotherapy.* 2009;1:539–47.
24. Goettel JA, Gandhi R, Kenison JE, Yeste A, Murugaiyan G, Sambanthamoorthy S, Griffith AE, Patel B, Shouval DS, Weiner HL, Snapper SB, Quintana FJ. AHR activation is protective against colitis driven by T cells in humanized mice. *Cell Rep.* 2016;17:1318–29.
25. Kaye J, Piryatinsky V, Birnberg T, Hingaly T, Raymond E, Kashi R, Amit-Romach E, Caballero IS, Towfic F, Ator MA, Rubinstein E, Laifenfeld D, Orbach A, Shinar D, Marantz Y, Grossman I, Knappertz V, Hayden MR, Laufer R. Laquinimod arrests experimental autoimmune encephalomyelitis by activating the aryl hydrocarbon receptor. *Proc Natl Acad Sci USA.* 2016;113:E6145–e6152.
26. Mezrich JD, Fechner JH, Zhang X, Johnson BP, Burlingham WJ, Bradfield CA. An interaction between kynurenine and the aryl hydrocarbon receptor can generate regulatory T cells. *J Immunol (Baltimore, MD : 1950).* 1950;185(2010):3190–8.
27. Singh NP, Singh UP, Singh B, Price RL, Nagarkatti M, Nagarkatti PS. Activation of aryl hydrocarbon receptor (AhR) leads to reciprocal epigenetic regulation of FoxP3 and IL-17 expression and amelioration of experimental colitis. *PLoS ONE.* 2011;6: e23522.
28. de Araújo EF, Loures FV, Preite NW, Feriotti C, Galdino NA, Costa TA, Calich VLG. AhR ligands modulate the differentiation of innate lymphoid cells and T helper cell subsets that control the severity of a pulmonary fungal infection. *Front Immunol.* 2021;12: 630938.
29. Huai W, Zhao R, Song H, Zhao J, Zhang L, Zhang L, Gao C, Han L, Zhao W. Aryl hydrocarbon receptor negatively regulates NLRP3 inflammasome activity by inhibiting NLRP3 transcription. *Nat Commun.* 2014;5:4738.
30. Masuda K, Kimura A, Hanieh H, Nguyen NT, Nakahama T, Chinen I, Ootoyo Y, Murotani T, Yamatodani A, Kishimoto T. Aryl hydrocarbon receptor negatively regulates LPS-induced IL-6 production through suppression of histamine production in macrophages. *Int Immunol.* 2011;23:637–45.
31. Rohlman D, Punj S, Pennington J, Bradford S, Kerkvliet NI. Suppression of acute graft-versus-host response by TCDD is independent of the CTLA-4-IFN-γ-IDO pathway. *Toxicol Sci Off J Soc Toxicol.* 2013;135:81–90.
32. Dant TA, Lin KL, Bruce DW, Montgomerly SA, Kolupaev OV, Bommasamy H, Bixby LM, Woosley JT, McKinnon KP, Gonzalez FJ, Blazar BR, Vincent BG, Coghill JM, Serody JS. T-cell expression of AhR inhibits the maintenance of pT(reg) cells in the gastrointestinal tract in acute GVHD. *Blood.* 2017;130:348–59.
33. Maurano MT, Wang H, John S, Shafer A, Canfield T, Lee K, Stamatoyannopoulos JA. Role of DNA methylation in modulating transcription factor occupancy. *Cell Rep.* 2015;12:1184–95.
34. Cong B, Zhang Q, Cao X. The function and regulation of TET2 in innate immunity and inflammation. *Protein Cell.* 2021;12:165–73.
35. Henry RA, Kuo YM, Andrews AJ. Differences in specificity and selectivity between CBP and p300 acetylation of histone H3 and H3/H4. *Biochemistry.* 2013;52:5746–59.
36. Gates LA, Shi J, Rohira AD, Feng Q, Zhu B, Bedford MT, Sagum CA, Jung SY, Qin J, Tsai MJ, Tsai SY, Li W, Foulds CE, O'Malley BW. Acetylation on histone H3 lysine 9 mediates a switch from transcription initiation to elongation. *J Biol Chem.* 2017;292:14456–72.
37. Karmodiya K, Krebs AR, Oulad-Abdelghani M, Kimura H, Tora L. H3K9 and H3K14 acetylation co-occur at many gene regulatory elements, while H3K14ac marks a subset of inactive inducible promoters in mouse embryonic stem cells. *BMC Genomics.* 2012;13:424.
38. Hua J, Chen Y, Fu B, Chen X, Xu XJ, Yang SH, Chen C, Xu YJ. Downregulation of p53 by insufficient CTCF in CD4(+) T cells is an important factor inducing acute graft-versus-host disease. *Front Immunol.* 2020;11: 568637.
39. Yamane H, Paul WE. Early signaling events that underlie fate decisions of naive CD4(+) T cells toward distinct T-helper cell subsets. *Immunol Rev.* 2013;252:12–23.
40. Roncarolo MG, Gregori S, Battaglia M, Bacchetta R, Fleischhauer K, Levings MK. Interleukin-10-secreting type 1 regulatory T cells in rodents and humans. *Immunol Rev.* 2006;212:28–50.
41. Apetoh L, Quintana FJ, Pot C, Joller N, Xiao S, Kumar D, Burns EJ, Sherr DH, Weiner HL, Kuchroo VK. The aryl hydrocarbon receptor interacts with c-Maf to promote the differentiation of type 1 regulatory T cells induced by IL-27. *Nat Immunol.* 2010;11:854–61.
42. Ehrlich AK, Kerkvliet NI. Is chronic AhR activation by rapidly metabolized ligands safe for the treatment of immune-mediated diseases? *Curr Opin Toxicol.* 2017;2:72–8.
43. Ehrlich AK, Pennington JM, Tilton S, Wang X, Marshall NB, Rohlman D, Funatake C, Punj S, O'Donnell E, Yu Z, Kolluri SK, Kerkvliet NI. AhR activation increases IL-2 production by alloreactive CD4(+) T cells initiating the differentiation of mucosal-homing Tim3(+) Lag3(+) Tr1 cells. *Eur J Immunol.* 2017;47:1989–2001.
44. Gandhi R, Kumar D, Burns EJ, Nadeau M, Dake B, Laroni A, Kozoriz D, Weiner HL, Quintana FJ. Activation of the aryl hydrocarbon receptor induces human type 1 regulatory T cell-like and Foxp3(+) regulatory T cells. *Nat Immunol.* 2010;11:846–53.
45. Singh NP, Singh UP, Rouse M, Zhang J, Chatterjee S, Nagarkatti PS, Nagarkatti M. Dietary indoles suppress delayed-type hypersensitivity by inducing a switch from proinflammatory Th17 cells to anti-inflammatory regulatory T cells through regulation of microRNA. *J Immunol (Baltimore MD).* 1950;196(2016):1108–22.
46. Ehrlich AK, Pennington JM, Bisson WH, Kolluri SK, Kerkvliet NI. TCDD, FICZ, and other high affinity AhR ligands dose-dependently determine the fate of CD4+ T cell differentiation. *Toxicol Sci Off J Soc Toxicol.* 2018;161:310–20.
47. Michonneau D, Latis E, Curis E, Dubouchet L, Ramamoorthy S, Ingram B, de Latour RP, Robin M, de Fontbrune FS, Chevret S, Rogge L, Socié G. Metabolomics analysis of human acute graft-versus-host disease reveals changes in host and microbiota-derived metabolites. *Nat Commun.* 2019;10:5695.

48. Kumari R, Palaniyandi S, Hildebrandt GC. Metabolic reprogramming—a new era how to prevent and treat graft versus host disease after allogeneic hematopoietic stem cell transplantation has begun. *Front Pharmacol.* 2020;11: 588449.
49. Xiao T, Wongtrakongate P, Trainor C, Felsenfeld G. CTCF recruits centromeric protein CENP-E to the pericentromeric/centromeric regions of chromosomes through unusual CTCF-binding sites. *Cell Rep.* 2015;12:1704–14.
50. Hou C, Zhao H, Tanimoto K, Dean A. CTCF-dependent enhancer-blocking by alternative chromatin loop formation. *Proc Natl Acad Sci U S A.* 2008;105:20398–403.
51. Ribeiro de Almeida C, Heath H, Krpic S, Dingjan GM, van Hamburg JP, Bergen I, van de Nobelen S, Sleutels F, Grosveld F, Galjart N, Hendriks RW. Critical role for the transcription regulator CCCTC-binding factor in the control of Th2 cytokine expression. *J Immunol (Baltimore MD).* 1950;182(2009):999–1010.
52. Kim LK, Esplugues E, Zorca CE, Parisi F, Kluger Y, Kim TH, Galjart NJ, Flavell RA. Oct-1 regulates IL-17 expression by directing interchromosomal associations in conjunction with CTCF in T cells. *Mol Cell.* 2014;54:56–66.
53. Liu Y, Wang L, Predina J, Han R, Beier UH, Wang LC, Kapoor V, Bhatti TR, Akimova T, Singhal S, Brindle PK, Cole PA, Albelda SM, Hancock WW. Inhibition of p300 impairs Foxp3⁺ T regulatory cell function and promotes antitumor immunity. *Nat Med.* 2013;19:1173–7.
54. Du T, Nagai Y, Xiao Y, Greene MI, Zhang H. Lysosome-dependent p300/FOXP3 degradation and limits Treg cell functions and enhances targeted therapy against cancers. *Exp Mol Pathol.* 2013;95:38–45.
55. Yang R, Qu C, Zhou Y, Konkel JE, Shi S, Liu Y, Chen C, Liu S, Liu D, Chen Y, Zandi E, Chen W, Zhou Y, Shi S. Hydrogen sulfide promotes Tet1- and Tet2-mediated Foxp3 demethylation to drive regulatory T cell differentiation and maintain immune homeostasis. *Immunity.* 2015;43:251–63.
56. Yue X, Lio CJ, Samaniego-Castruita D, Li X, Rao A. Loss of TET2 and TET3 in regulatory T cells unleashes effector function. *Nat Commun.* 2019;10:2011.
57. Xu YJ, Chen FP, Chen Y, Fu B, Liu EY, Zou L, Liu LX. A possible reason to induce acute graft-vs.-host disease after hematopoietic stem cell transplantation: lack of sirtuin-1 in CD4(+) T cells. *Front Immunol.* 2018;9:3078.
58. Glucksberg H, Storb R, Fefer A, Buckner CD, Neiman PE, Clift RA, Lerner KG, Thomas ED. Clinical manifestations of graft-versus-host disease in human recipients of marrow from HL-A-matched sibling donors. *Transplantation.* 1974;18:295–304.
59. Przepiorka D, Weisdorf D, Martin P, Klingemann HG, Beatty P, Hovs J, Thomas ED. Consensus conference on acute GVHD grading. *Bone Marrow Transplant.* 1994;15:825–8.
60. Abron JD, Singh NP, Mishra MK, Price RL, Nagarkatti M, Nagarkatti PS, Singh UP. An endogenous aryl hydrocarbon receptor ligand, ITE, induces regulatory T cells and ameliorates experimental colitis. *Am J Physiol Gastrointest Liver Physiol.* 2018;315:G220-g230.

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